

Neurotrophins and Their Receptors—Current Concepts and Implications for Neurologic Disease

FERNETTE F. EIDE,*† DANIEL H. LOWENSTEIN,*‡ AND LOUIS F. REICHARDT†§

Departments of *Neurology and †Physiology, ‡Epilepsy Research Laboratory, and §Howard Hughes Medical Institute, University of California at San Francisco, California 94143-0724

Nerve growth factor, brain-derived neurotrophic factor, neurotrophin-3, and neurotrophin-4/5 are members of a family of proteins (the neurotrophins) that promote the differentiation, growth, and survival of peripheral and central nervous system neurons. Recently, the *trk* family of proto-oncogenes has been found to encode signal-transducing receptors for these growth factors. This discovery has important implications for our understanding of the normal function of these factors in the developing and adult nervous system. In this review, we highlight recent advances in neurotrophin research and discuss their relevance to neurologic disease. © 1993 Academic Press, Inc.

INTRODUCTION

The neurotrophins are factors that regulate the growth and survival of selected peripheral and central nervous system neurons. As *trophic* (survival-promoting) and *tropic* (directing axon growth) factors, they have been investigated for use in treatment of several types of neurodegenerative disease, peripheral neuropathy, and nervous system cancer. In this paper, we discuss recent advances in neurotrophin research and review their therapeutic potential for human disease.

NERVE GROWTH FACTOR

Nerve growth factor (NGF) was discovered as a sarcoma-derived substance that stimulated the outgrowth of dorsal root ganglion neurons (118). It is now recognized to be a survival and differentiating factor for neural crest sensory neurons and sympathetic and basal forebrain cholinergic neurons (106, 142, 201). In addition, NGF is a proliferative factor for striatal neuroepithelial precursors (27) and possibly an activity-dependent organizer of the visual cortex (21, 39). As a target-derived trophic factor, NGF is released at innervation sites, where it is internalized at nerve terminals and retrogradely transported back to the cell body (77, 110, 178). By controlling survival at the time of target inner-

vation, NGF can help regulate the size and connectivity of neuronal populations.

Biologically active NGF consists of two noncovalently linked peptide chains (5, 156). Each chain is comprised of 118 amino acids that includes 6 strictly conserved cysteines (4). In its crystal structure, the NGF dimer contains three antiparallel pairs of β -strands (125). Variable residues located in three hairpin loops are believed to contribute to different biological properties observed among members of the nerve growth factor family.

THE NEUROTROPHIN GROWTH FACTOR FAMILY

In 1982, Barde and colleagues isolated a trophic factor from pig brain that promoted the outgrowth of spinal sensory neurons (14). That protein, known as brain-derived neurotrophic factor (BDNF) was cloned in 1989 and found to share close sequence homology with NGF (50% sequence identity) (118). BDNF supports the survival and outgrowth of neural crest-derived and placode-derived sensory neurons, parasympathetic nodose ganglion neurons, dopaminergic striatal neurons, basal forebrain cholinergic neurons, retinal ganglion neurons, and cranial and spinal motoneurons, but not sympathetic neurons (118, 150, 163, 176, 215). In addition to its survival and outgrowth-promoting activities, BDNF may also play a role in the cell fate determination of primary sensory neurons (180) and the activity-dependent organization of visual cortex (26). Across species lines, BDNF appears to have a higher degree of sequence conservation than nerve growth factor (65).

In 1991, neurotrophin-3 (NT-3) and neurotrophin-4/5 (NT-4/5) were cloned using the polymerase chain reaction and sequence identities shared between BDNF and NGF (23, 44, 70, 79, 95, 96, 127, 164). Mature NT-3 shared 50 identical amino acids in common with NGF and BDNF, including 6 cysteine residues. Across species lines, NT-3 also shows remarkable conservation—the sequences for NT-3 are identical in rats, mice, and humans (96). Neurotrophin-4 and neurotrophin-5 were initially thought to be distinct growth factors (23, 70). However, recent evidence suggests they are homologues of a

TABLE 1

Neuronal Selectivities of Neurotrophin Growth Factors			
Neurotrophin	Responsive neurons	References	
NGF	Sympathetic neurons Neural crest sensory neurons	(119, 142)	
			PNS
CNS	Basal forebrain cholinergic neurons Striatal cholinergic neurons Cerebellar Purkinje cells	(30, 140, 141)	
BDNF	Placode-derived sensory neurons Neural crest sensory neurons Nodose ganglion neurons	(22, 36, 150, 215)	
			PNS
CNS	Spinal motoneurons Basal forebrain cholinergic neurons Proprioceptive trigeminal neurons Substantia nigra dopaminergic neurons Retinal ganglion neurons Facial motoneurons	(36, 86, 163, 176, 206)	
NT-3	Sympathetic neurons Sensory neurons	(51, 79, 127, 164)	
			PNS
CNS	Basal forebrain cholinergic neurons Locus coeruleus		
NT-4/5	Sympathetic neurons Dorsal root ganglion neurons Nodose ganglion neurons	(23, 51, 70)	
			PNS
CNS	Basal forebrain cholinergic neurons Locus coeruleus		

single factor expressed in different animal species (88). NT-4/5 appears to be less highly conserved across species lines, although it shares over 50% sequence homology with NGF, BDNF, and NT-3 (23, 69). All neurotrophins can support the growth and survival of subsets of dorsal root ganglion neurons; however, they differ in their sites of synthesis, developmental patterns of expression, and neuronal targets (see Table 1).

THE NEUROTROPHIN RECEPTORS

High- and Low-Affinity Receptors

Receptors for neurotrophin growth factors consist of two general types—the high-affinity or “slow-dissociat-

ing” receptors and the low-affinity or “fast-dissociating” receptors. High-affinity receptors ($K_d \sim 10^{-11}$) distinguish between members of the neurotrophin family and appear to transduce most, if not all, of these factors' biological effects (83, 169, 193). Low-affinity receptors bind all neurotrophins with comparable low affinity ($K_d \sim 10^{-9}$) (22) and have a largely undefined role in the signaling process (discussed further below). High-affinity receptors have molecular weights of 140 kDa (135) and low-affinity receptors have molecular weights of 75 kDa (93). High- and low-affinity receptors can also be distinguished on the basis of their ligand specificity, dissociation kinetics, protease sensitivity, and detergent solubility (24, 115, 193, 211).

The first indication that high-affinity receptors were responsible for most of the neurotrophins' biological effects came from the *in vitro* observation that the concentration of NGF required to elicit a biological response was consistent with high-affinity binding (66). Subsequent *in situ* NGF-binding studies showed that the biological responsiveness of a tissue correlated with the expression of high- but not low-affinity receptors (162). The low-affinity receptor is found in neuronal and nonneuronal tissues not clearly responsive to NGF (214).

Trk Proto-oncogenes Encode Receptors for Neurotrophin Signal Transduction

In 1991, investigators discovered that at least one subunit of the high-affinity receptor for NGF was encoded by the *trk A* proto-oncogene (98, 99, 103). *Trk A* was originally discovered in tissue obtained from a human colon carcinoma biopsy (130). In its malignant form, the catalytic and transmembrane domains of *trk A* were fused with nonmuscle tropomyosin, resulting in the constitutive activation of *trk A* (33). The *trk A* proto-oncogene showed preferential expression in the nervous system (129). However, no link to NGF was made until investigators demonstrated that nonneuronal cells transfected with *trk A* acquired NGF-binding sites and activated intracellular signal transduction pathways (92, 98, 102). Fibroblasts transfected with *trk A* bound ^{125}I -NGF with high affinity and became morphologically transformed (92), and *Xenopus* oocytes transfected with *trk A* underwent maturational changes following exposure to NGF (146). *Trk* receptors have now been detected in every major population of neurons known to respond to NGF (82, 129). Shooter and colleagues have also demonstrated that *trk A* receptors possess kinetic and chemical properties characteristic of slow-dissociating receptors (136).

It is now known that the *trk* family of proto-oncogenes encodes three closely related tyrosine kinases—*trk A*, *trk B*, and *trk C* (14, 131, 138). The *trk* proteins have a tripartite structure consisting of an extracellular

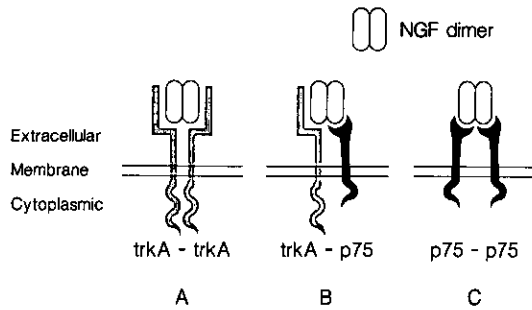


FIG. 1. Receptor models for nerve growth factor signal transduction. Three receptor models have been proposed for nerve growth factor signal transduction: (A) trk A-trk A homodimer; (B) trk A-p75 (low-affinity receptor) heterodimer; (C) p75-p75 homodimer.

domain for ligand recognition, a single transmembrane domain, and a cytoplasmic tail (containing the tyrosine kinase) for initiating the signaling cascade. The extracellular portion includes two immunoglobulin-like modules that may form a pocket for growth factor binding (174). Tyrosine kinase receptors are believed to be activated by a process of allosteric dimerization (207). In this model, growth factor binding causes tyrosine kinase receptors to pair (or form oligomers). When the cytoplasmic domains are brought into close proximity, they activate kinase domains. Figure 1 shows three hypothetical receptor dimers (trk A-trk A, trk A-p75, p75-p75) postulated in the signal transduction of NGF. The trk A receptor appears to be an essential mediator of the signaling response to NGF (87, 103, 204); the role of p75 is less clear (see discussion below).

A puzzling feature of neurotrophin signal transduction is the degree of redundancy that exists between the neurotrophin growth factors and their receptors. Trk A encodes a signal-transducing receptor for NGF (and NT-3 at lower efficacy) (31, 98, 99, 103, 146). Trk B is a signal-transducing receptor for BDNF (and NT-3 at lower efficacy) (186, 188). Trk C is a signal-transducing receptor for NT-3 (114). NT-4/5 is reportedly able to bind to and activate either trk A or trk B (23, 88) (Table 2). Because of restrictions in the timing and distribution of growth factors (and their receptors), functional redundancy may be rare *in vivo*. However, the presence of redundancy in other growth factor systems (200) suggests that the overlaps could have some biological signif-

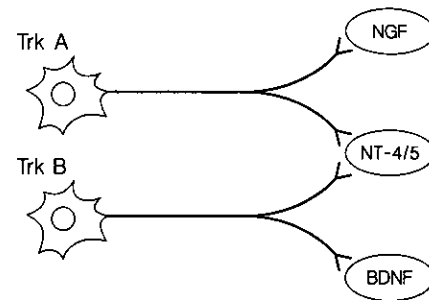


FIG. 2. Possible role for neurotrophin redundancy in the development of synaptic connectivity. The ability of a trk receptor to respond to multiple neurotrophins could have a role in the establishment of appropriate neuronal connections. For instance, a target tissue expressing NT-4/5 can support two distinct populations of neurons (one expressing trk A, the other expressing trk B). Although both trk A- and trk B-expressing neurons converge onto the NT-4/5 target, they are also able to form other divergent projections to NGF- or BDNF-expressing tissues. The redundancy in neurotrophins would also increase the availability of trophic support to a particular group of neurons.

icance. In some instances, differences in the temporal expression of factors (or their receptors) could allow a single factor to have multiple functions throughout the course of development. In addition, during embryonic development, the redundancy between neurotrophins and their receptors could be useful in the establishment of appropriate neuronal connections. As suggested in Fig. 2, the responsiveness of a single receptor to multiple neurotrophins would allow a tissue expressing different neurotrophins to establish divergent projections. By allowing a single factor to activate multiple receptors, different neuronal groups could share in trophic support. Also, during development, neurons could utilize different neurotrophins expressed along their pathways of growth.

Splice Variants

Splice variants of trk receptors represent another puzzling feature of neurotrophin signal transduction. Alternative transcripts for trk A, trk B, and trk C have been identified in the rat, mouse, and chick nervous systems (15, 59, 102, 138). The patterns of sequence divergence suggests that variants are generated by a differential splicing of alternative exons. The variants have included receptors that lack functional kinase sequences (truncated receptor) and receptors with either alternative 5' terminals or kinase insertions. The truncated receptor appears identical to the full-length receptor in its extracellular and transmembrane domains, but lacks a cytoplasmic tyrosine kinase sequence. This receptor would bind neurotrophins normally, but fail to activate intracellular signal transduction pathways. Several putative functions for the truncated receptors have been proposed: (1) a role in ligand presentation (e.g.,

TABLE 2

Redundancy of Neurotrophins and Their Receptors

Receptor	Neurotrophin
Trk A	NGF, NT-3, NT-4/5
Trk B	BDNF, NT-3, NT-4/5
Trk C	NT-3

assisting in the presentation of neurotrophins to full-length receptors), (2) an activator of alternate signaling pathways (activating distinct signaling pathways from full-length receptors by stimulating cytoplasmic protein kinases), (3) a modulator of neurotrophin responsiveness (e.g., sequestering catalytic full-length receptors), and (4) a mediator of cell-cell interactions (e.g., having a role in axon guidance). Recently, Beck *et al.* found that truncated trk B receptors were induced in hippocampal pathways undergoing active synaptic reorganization (17). This group found that truncated trk B receptors were induced in reorganizing commissural/association pathways 6–14 days following perforant path-fimbria-fornix transections. In unlesioned mice, truncated trk B receptors have been found in choroid plexus and ependyma, consistent with clearance or active transport functions across the blood brain barrier (102).

p75: The Low-Affinity Receptor

When p75 was first isolated (93, 159), it was thought to be the signal-transducing receptor for NGF. It was named the low-affinity nerve growth factor receptor; however, it was later found to bind all neurotrophins with comparable affinity (70, 186, 188). p75 has a tripartite structure consisting of four cysteine-rich extracellular domains, a short serine- and threonine-rich spacer, and a cytoplasmic domain. The transmembrane and cytoplasmic domains of p75 are highly conserved between species, but share no known homology to signal-transducing proteins. p75's predicted tertiary structure is similar to that of mastoparan, an activator of GTP-binding regulatory proteins (46, 78), leading investigators to hypothesize that p75 may activate G proteins in its signal transduction pathway (16). The cysteine-rich extracellular domain of p75 shares sequence homology to genes encoding the receptors for tumor necrosis factor (184), T cell activation antigen OX-40 (128), B cell antigen CD40 (189), and Fas, a cell surface antigen that mediates apoptosis (90).

Although p75 is expressed in every neuronal tissue known to respond to NGF (43, 160), its role in the signaling process remains controversial. In membrane preparations, Chao and colleagues have reported that high-affinity NGF-binding sites were generated only if trk A was coexpressed with p75 (76). If membranes containing p75 and trk A were fused, high-affinity binding sites were generated *in vitro* (98). On the other hand, in whole cell preparations, Klein *et al.* (103) found that the expression of trk A (in the absence of p75) generated both high and low affinity sites for NGF. Chao and colleagues (216) have also reported that receptor chimeras, consisting of the extracellular domain of the epidermal growth factor receptor fused to the intracellular and transmembrane domains of p75, transduce NGF-like

responses upon stimulation with epidermal growth factor. However, these results contradict the observation that BDNF (a ligand that binds p75 with similar affinity to NGF) fails to promote neurite outgrowth in p75-containing PC12 cells. In PC12 cells, transfection with trk B confers responsiveness to BDNF (186). In other studies, Chao and colleagues have reported that overexpression of the cytoplasmic domain of p75 in PC12 cells down-regulates trk A kinase activity (19). Despite the loss of high-affinity sites, transfected cells appeared to differentiate normally in response to NGF.

Persson and colleagues have examined the potential contribution of p75 by designing a mutant NGF molecule unable to bind p75 (87). This mutant retained its binding site for trk A and stimulated neurite outgrowth and dorsal root ganglion survival at physiological concentrations. Members of our group have also shown that polyclonal antibodies that block NGF-p75 binding have no effect on NGF's trophic effects (204). Recently, Barbacid *et al.* have reported that kinase-deficient trks can act as dominant negative mutants in NIH3T3 cells stably transfected with trk A (92). When kinase-deficient mutants are introduced into transfected cells, they form noncatalytic dimers with full-length receptors, blocking NGF-induced morphological transformation. The expression of p75 in these cells had no effect on high-affinity binding or transformation events.

Recently, Wheeler and Bothwell questioned whether functional assays such as neurite outgrowth or neuronal survival were adequate to detect biological functions of p75 (206). From *in situ* hybridization studies, they found that p75 expression in the developing limb bud, kidney, lung, and testes correlated better with sites of tissue induction than with innervation. In the developing limb bud, for example, NGF and p75 were expressed at sites of limb attachment and in the mesenchyme adjacent to the apical ectodermal ridge, rather than at sites of innervation. In the developing kidney, p75 was expressed in polarizing epithelium during glomerular morphogenesis. More direct evidence for p75's role in kidney morphogenesis is supported by the observation that antisense p75 oligonucleotides block glomerular formation in kidney explants (167). Recently Jaenisch and colleagues generated transgenic mice deficient in the gene encoding p75 (117). No kidney abnormalities in these animals were seen. However, sensory abnormalities and trophic ulcers developed by age 4 months. Immunohistochemical analyses of footpad skin revealed decreased innervation by calcitonin gene-related peptide- and substance P-immunoreactive fibers. When the human p75 gene was crossed into mutant animals, the phenotype was reversed. Although these findings support a role for p75 in the normal development and function of peripheral sensory neurons, the mutant phenotype could occur as a result of an indirect effect on Schwann cells. Schwann cells express p75 during and just prior to

cutaneous innervation (212). Their expression of p75 receptors may function in growth and guidance of developing axons (166).

IMPLICATIONS FOR NEUROLOGIC DISEASE

Alzheimer's Disease

Much of the initial clinical interest in NGF centered on its therapeutic potential in the treatment of Alzheimer's disease (AD). In AD, severe neuronal loss, neurofibrillary tangles, and neuritic plaques disrupt learning and memory pathways, resulting in the severe intellectual decline seen in AD (85). The cholinergic basal forebrain and septum (areas implicated in memory and learning) are severely affected by neuronal loss (37, 208). Since basal forebrain and septal cholinergic neurons are major central nervous system targets for NGF, NGF has been actively investigated for potential use as a therapy for AD. Over the past decade, several experimental paradigms have been used to address this potential application: (1) fimbria-fornix transection, (2) retrograde lesions of nucleus basalis magnocellularis (nbM), (3) the use of memory-impaired aged rats, and (4) a transplantation model of the trisomy 16 mouse.

If mammalian fimbria-fornix pathways are transected with a knife, 65–90% of medial septal neurons develop retrograde degenerative changes and die (10). However, if intraventricular NGF is administered at the time of lesioning, most cholinergic septal/diagonal band neurons will survive (54, 73, 108, 111, 199). Even with delayed administration of NGF, partial recovery of cholinergic neurons is seen (69).

Intraventricular NGF can also prevent lesion-induced atrophy in the nbM (71, 190). Following retrograde injury to the nbM, no decrease in neuronal number is seen, although choline acetyltransferase (ChAT) activity falls. If animals are treated with intraventricular NGF, levels of ChAT in the nucleus basalis and cortical areas spared by the lesion return to normal. NGF treatment has also been reported to ameliorate memory and learning deficits caused by fimbria-fornix or nucleus basalis lesions. However, these results are difficult to interpret because memory deficits created by septal lesions are exacerbated by NGF (152).

In experimental models of age-related memory impairment, NGF infusion improves the behavioral performance of impaired animals (49, 53). When aged rats were segregated on the basis of their performance on spatial memory tasks, impaired animals were found to have smaller cholinergic basal forebrain neurons (48). When impaired animals received intraventricular infusions of NGF, their basal forebrain neurons increased in size and their performance on the swimming maze improved (49). Using immunocytochemical methods, Koh and Loy (107) have reported an age-related decline in

p75 immunoreactivity in the rat basal forebrain. In human tissue, Hefti and Mash (74) have also found age-related decreases in the NGF receptor. To date, one study has reported a decline in NGF protein and mRNA in the brains of aged rats (116).

Recently, the trisomy 16 (Ts16) mouse has provided a genetic model of basal forebrain abnormalities occurring in Down syndrome and Alzheimer's disease. A striking genetic homology exists between the distal long arm of human chromosome 21 and the distal end of mouse chromosome 16 (MMU16) (28, 34). At least six genes are shared between MMU16 and human chromosome 21, including the genes encoding amyloid precursor protein, superoxide dismutase, and markers linked to a familial form of AD. Ts16 mice share many of the phenotypic features of human Down syndrome, including basal forebrain cholinergic atrophy (34, 100, 195). Since trisomy 16 mice die at birth, experiments must be based on either *in vitro* or transplantation approaches.

The basal forebrains of Ts16 mice have fewer cholinergic neurons than euploid littermates and remaining neurons are small with short processes lacking varicosities (100, 195). If Ts16 neurons are cultured in the presence of NGF, they hypertrophy, extend processes (with varicosities), and increase their ChAT activity (32). In a transplantation model of the Ts16 mouse, Ts16 neurons integrate into normal mouse hippocampus and initially appear indistinguishable from euploid neurons. Six months later, however, Ts16 neurons atrophy and lower levels of ChAT activity are seen [81; D. M. Holtzman and W. C. Mobley, unpublished observations]. If NGF is administered before the genetically programmed atrophy begins (at 5½ months), basal forebrain cholinergic neurons retain their normal appearance and ChAT levels remain high.

Although NGF would appear to have therapeutic potential in the treatment of AD, critics have warned that available NGF models (fimbria-fornix transection, nbM lesioning, trisomy 16 mouse) do not replicate the essential findings of Alzheimer's disease. In fact, none of the animals in these paradigms develop β -amyloid plaques, neurofibrillary tangles, or amyloid angiopathy. Because point mutations in the β A4-amyloid precursor protein (β -APP) gene have been found to be responsible for certain kindreds of familial early-onset AD (61, 145, 202), it is important to consider the effect of NGF on normal and abnormal β -amyloid processes.

In hamster basal forebrain and SH-SY5Y neuroblastoma cells, NGF increases β -APP mRNA expression (109, 139). The β -APP gene generates at least five alternative translation products that include APP₆₉₅, APP₇₅₁, and APP₇₇₀ (64, 97, 101, 158, 197). APP₇₅₁ and APP₇₇₀ are expressed predominantly in peripheral tissues; APP₆₉₅ is expressed in developing and adult brain (101, 109, 147, 158, 197). In PC12 cells, NGF increases the expression of APP₆₉₅, decreases the expression of

APP₇₇₀, and increases β -APP release (161, 175, 185). Although the significance of these fluctuations is unclear regarding the pathogenesis of Alzheimer's disease, Yoshikawa *et al.* have recently shown that overexpression of APP₆₉₅ or APP₇₇₀ in P19 embryonal carcinoma cells results in the death of postmitotic neurons (218). They hypothesized that the overproduction of β -APP could overwhelm endogenous processing, leading to a generation of amyloidogenic fragments and the death of postmitotic neurons derived from these cells. β -APP expression has also been implicated in the aberrant sprouting responses seen in AD (133). In either instance the administration of NGF in this setting could exacerbate the pathological cascade of AD. In fact, Yankner *et al.* (1990) have reported that NGF treatment greatly increases the neurotoxicity of β -amyloid (10^5 -fold) *in vitro* (217). β -amyloid also increased NGF receptor immunoreactivity in this study, leading investigators to hypothesize that pathological accumulations of β -amyloid could activate a positive feedback loop between β -amyloid and NGF, accelerating the death of NGF-responsive neurons.

Because many neuronal groups affected in AD are unresponsive to NGF, a primary defect in NGF-mediated pathways is not believed to underlie AD pathogenesis. Trk receptor expression has not been systematically examined in Alzheimer tissue. However, normal levels of NGF and p75 mRNA have been seen in AD brains at autopsy (62, 74). BDNF, NT-3, and NT-4/5 have recently been found to be survival-promoting factors for cholinergic basal forebrain neurons (51, 106). Their trophic effects *in vivo* are currently under investigation. Thus far, Knusel *et al.* (105) have demonstrated a slight, but significant, effect for BDNF on axotomized cholinergic septal and diagonal band neurons. BDNF appeared to rescue a portion of lesioned neurons, whereas NGF treatment rescued the entire population. Phillips *et al.* (155) have reported that the dentate gyri of AD patients express significantly lower levels of BDNF mRNA than those of nondemented controls. However, it is likely that these changes reflect secondary processes in the disease.

Parkinson's Disease

Parkinson's disease (PD) is a debilitating movement disorder caused by the selective death of dopamine-containing substantia nigra neurons (2). Until recently, acidic and basic fibroblast growth factor (aFGF, bFGF) were the only trophic factors known to promote the growth and survival of dopaminergic midbrain cells.

aFGF and bFGF are tyrosine kinase-mediated growth factors unrelated to the neurotrophin growth factor family. Other members of this family include INT2, HST, FGF5, FGF6, and KGF (20). If fetal dopamine-containing mesencephalic neurons are cultured in the presence

of aFGF or bFGF, they show increased survival and neurite outgrowth (47). In 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-treated mice, both aFGF and bFGF appear to prevent a loss of tyrosine hydroxylase (TH)-positive striatal neurons (151). However, a potential problem with mouse MPTP paradigms is that their dopaminergic neurons do not degenerate, despite the loss of TH immunoreactivity, and spontaneous regeneration is sometimes seen (179). In other studies, aFGF has been seen to have no effect on aged MPTP-exposed mice (35), and bFGF had no protective effect *in vitro* (72). Recently Engele and Bohn (41) have presented evidence that mesencephalic glia mediate the FGF's trophic effects on dopaminergic midbrain. Coincident with a rise in TH+ cells (in response to aFGF or bFGF) was a tripling of mesencephalic glia. If the glial proliferation was blocked by mitotic inhibitors, no neurotrophic effects were seen. Also, conditioned media from glial cultures supported dopaminergic neuron survival. In fact, the potent mitogenic effects of FGF's are likely to exclude their use in the treatment of neurodegenerative disease. bFGF stimulates the proliferation of human glioma cells (205) and elevated levels of bFGF have been reported in human glioma cell lines (168, 196).

BDNF has also been found to increase the survival, outgrowth, and dopamine uptake of embryonic dopaminergic midbrain neurons (86, 106). *In vitro*, BDNF is able to protect neurons from MPTP- or 6-hydroxydopamine-induced injury (106, 187); however, *in vivo* experiments with MPTP have not yet been reported. Hefti and colleagues have recently found no effect for BDNF on partially transected substantia nigra neurons (105). In other experiments nigral injections of BDNF have been found to stimulate rotational behavior and increase striatal levels of dopamine metabolites (3). To date, p75 and trk B have not been found in the substantia nigra (104, 157). An indirect action of BDNF on dopaminergic neurons cannot be excluded.

In adrenal transplantation approaches to PD, NGF has shown promise in increasing the process formation, differentiation, and survival of chromaffin grafts (40). In rodent models of Parkinson's disease, NGF treatment resulted in a longer graft survival and better functioning for the life of the animal (181, 192). Unfortunately, results in monkeys and humans have been disappointing. In man, adrenal graft survival has been poor and behavioral recovery slight (84, 153). Numerous groups have failed to reproduce the striking surgical successes of Madrazo and colleagues (1, 63, 126). In a single human case, murine NGF was administered intraputaminally to adrenal transplant recipient (148). NGF caused no apparent deleterious effects and appeared to prolong graft survival.

Amyotrophic Lateral Sclerosis

Amyotrophic lateral sclerosis is a disease in which cranial and spinal motoneurons degenerate, resulting in a

debilitating syndrome of generalized weakness, wasting, fasciculation, and pyramidal tract dysfunction (210). The treatment is supportive, and most patients die within 4 years (25).

In 1990, Thoenen and colleagues discovered that ciliary neurotrophic factor (CNTF), a growth factor unrelated to the neurotrophins, could support the survival of cultured spinal motoneurons (9, 13). Although CNTF is not expressed in muscle (191), it rescues motoneurons from developmental or axotomy-induced death (150, 177). More recently, CNTF's trophic effects have been demonstrated in genetic animal models of motoneuron death (the wobbler, pmn, and mnd mouse) (75, 80, 176). In these studies, CNTF prevented motoneuron loss and slowed the progression of disease.

CNTF belongs to a class of neurotrophic molecules that transduce their biological signals on cytokine-like receptors (38, 113). Unlike the neurotrophins, CNTF lacks a signal sequence and glycosylation sites and is not secreted. Because rising levels of Schwann cell CNTF have correlated with an age-related increase in the survival of lesioned motoneurons (191), some investigators have hypothesized that CNTF acts as a "lesion factor" that is released under pathological conditions. Because of CNTF's promising performance in animal studies, it has entered Phase II trials for use in the treatment of human motoneuron disease.

In 1992, BDNF was also found to be a trophic factor for cranial and spinal motoneurons (150, 176, 215). In the neonatal rat, local applications of BDNF prevented the axotomy-induced death of spinal and facial motoneurons (176, 215). In the developing chick, BDNF prevented naturally-occurring motoneuron death (150). Because BDNF is expressed by developing muscle and retrogradely transported in spinal motoneurons (170, 215), it appears to be a normal regulator of motoneuron growth. Interestingly, Barbacid and colleagues have found that mice carrying a targeted mutation of *trk B* have obvious motor and sensory deficits (M. Barbacid, personal communication). Mice homozygous for the mutation have been found to have fewer facial and spinal motoneurons than wild-type or heterozygote siblings.

Peripheral Neuropathy

Since NGF is required for the normal survival of adult sympathetic and sensory neurons, it has also been investigated for use as a therapeutic agent in toxic and diabetic peripheral neuropathies. Disabling sensory neuropathies frequently limit the dosing of chemotherapeutic agents in cancer therapy. Taxol, a chemotherapeutic agent used in the treatment of melanoma and ovarian cancer (182), can disrupt axonal transport in peripheral neurons. This impairs the trophic supply of factors like NGF and causes painful sensory symptoms (123). When

dorsal root ganglion explants are exposed to taxol *in vitro*, they accumulate abnormal aggregations of microtubules, disrupting the cytoskeleton and resulting in cell death (172). If NGF is added to culture media, >90% of dorsal root ganglion neurons are rescued (154). In 1991, Apfel *et al.* developed an animal model of taxol neuropathy that reproduced changes in pain threshold, compound nerve action potentials, and dorsal root ganglion levels of substance P (8). When NGF was administered with taxol injections in this model, all three parameters improved. Similar findings have been reported for NGF treatment in a cisplatin model (7). However, two of the parameters used in this study (nerve conduction velocity, calcitonin gene-related peptide levels) have made the significance of these findings unclear. Again, it seems likely that other neurotrophins may be useful in maintaining sensory innervation and functions.

NGF has also been considered as a therapeutic factor in diabetic neuropathy because peripheral sensory and sympathetic neurons are affected early in the disease (45). In a streptozotocin model of diabetes, several groups have found decreased retrograde axonal transport of ¹²⁵I-NGF (137, 173). When NGF treatment was tested in this paradigm, it prevented some behavioral and biochemical manifestations of small fiber neuropathy, but failed to prevent nerve conduction velocity slowing (6).

Nervous System Cancer

The relationship between neurotrophin growth factors and primitive neuroectodermal tumors (PNETs) has been studied because members of the neurotrophin growth factor family affect the survival, proliferation, and differentiation of immature neural crest cells (121, 127). PNET tumors share a number of histological and functional similarities to immature neural crest cells, and it has been hypothesized that these cancers arise as a defect in normal differentiation (18). Neuroblastomas, medulloblastomas, ganglioneuromas, and Ewing's sarcomas have all been found to express the low-affinity neurotrophin receptor (165). Recently, Nakagawa *et al.* (144) noted an inverse relationship between the expression of *trk A* and *n-myc* amplification in a large number of human neuroblastoma cell lines. High levels of *trk A* expression correlated best with highly differentiated neuroblastomas and a favorable clinical prognosis; low levels of *trk A* correlated with poor differentiation and a poor prognosis. Tumors expressing high levels of *trk A* were more likely to differentiate, regress spontaneously, and respond to chemotherapy. Interestingly, treatment by NGF can cause responsive tumor cell lines to grow more slowly and acquire differentiated morphologies (91, 132, 134, 203). *In vivo*, NGF has been shown to slow the growth of intracerebrally implanted gliomas (203) and subcutaneous neurinomas

(213). In 1970, three children with disseminated neuroblastoma received five to eight intramuscular injections of murine NGF (10 mg/day) (112). Although no clear beneficial effects were noted, the treatment was tolerated with minimal side effects.

Epilepsy

The role of neurotrophins in epileptogenesis has also emerged as a promising area of scientific investigation. In the past few years, there has been a growing body of evidence that a reorganization of neural networks is an essential feature of certain forms of epilepsy. A consistent pathological finding in the temporal lobes of patients with intractable epilepsy is the mossy fiber sprouting of dentate granule cells (11, 194). In fact, quantitative histological and ultrastructural studies show that the extent of mossy fiber reorganization correlates with the occurrence of seizures. Although it is unclear whether this synaptic reorganization is maladaptive (60, 160, 198) or adaptive (183), it is an active area of investigation because it represents a potentially novel site for therapeutic intervention in epileptogenesis.

The association of abnormal electrical activity with synaptic reorganization led a number of investigators to study changes in neurotrophin expression following experimentally induced seizures. In 1989, Gall and Isackson (56) demonstrated that limbic seizures increased the expression of NGF mRNA in hippocampal pathways. Since then, seizures induced by intraventricular or intraperitoneal kainic acid have also been shown to increase NGF expression (42, 58, 89, 219). An increase in NGF expression can even be detected after a single epileptiform hippocampal discharge (57). More recently, BDNF expression has also been found to increase following the induction of limbic seizures (12, 89). No change in NT-3 expression was seen (42).

In vitro studies have confirmed that a rapid increase in neurotrophin synthesis can be induced by increased neuronal activity. Depolarizing stimuli such as high extracellular potassium, veratridine, or treatment with glutamate agonists cause prompt rises in NGF or BDNF expression in hippocampal neurons (124, 219). It is likely that the abnormal electrical activity that occurs in human epilepsy results in changes in neurotrophin expression. Although the functional significance of these changes remains unproven, it is interesting that Funi-bashi *et al.* (52) found that centrally administered antiserum to NGF delayed the onset of amygdaloid kindling.

FUTURE PROSPECTS

The past 5 years have seen great advances in our understanding of the normal biology and therapeutic potential of neurotrophin growth factors and their recep-

tors. Since 1989, BDNF, NT-3, and NT-4/5 have been sequenced, trk proto-oncogenes have been found to encode neurotrophin signal transducing receptors, and the three-dimensional structure of NGF has been solved. Existing data have suggested that neurotrophin growth factors have therapeutic potential in the treatment of certain types of neurodegenerative disease, peripheral neuropathy, cancer, and epilepsy. With technical advances in gene and drug delivery, neural transplantation, and rational drug design, practical therapeutic hurdles are likely to be overcome in the near future.

ACKNOWLEDGMENTS

We thank Drs. David Holtzman, William Mobley, and Brock Eide for their helpful discussions and critical comments on this manuscript. We would also like to thank Drs. Mariano Barbacid, David Holtzman, and William Mobley for allowing us to communicate work prior to its publication. This work was supported in part by grants from the American Epilepsy Society, NIH K11AG0056801, MH48200, and NS01424. L. F. Reichardt is an investigator for the Howard Hughes Medical Institute.

REFERENCES

1. AHLSSKOG, J. E., P. J. KELLY, J. A. VAN EERDEN, S. L. STODDARD, G. M. TYCE, A. J. WINDEBANK, P. A. BAILEY, G. N. BELL, M. D. BLEXRUD, AND S. W. CARMICHAEL. 1990. Adrenal medullary transplantation into the brain for treatment of Parkinson's disease: clinical outcome and neurochemical studies. *Mayo Clin. Proc.* **65**: 305-328.
2. ALBIN, R. L., A. B. YOUNG, AND J. B. PENNEY. 1989. The functional anatomy of basal ganglia disorders. *Trends Neurosci.* **12**: 366-375.
3. ALTAR, C. A., C. B. BOYLAN, C. JACKSON, S. HERSHENSON, J. MILLER, S. J. WIEGAND, R. M. LINDSAY, AND C. HYMAN. 1992. Brain-derived neurotrophic factor augments rotational behavior and nigrostriatal dopamine turnover *in vivo*. *Proc. Natl. Acad. Sci. USA* **89**: 11347-11351.
4. ANGELETTI, R. H., AND R. A. BRADSHAW. 1971. Nerve growth factor from mouse submaxillary gland: amino acid sequence. *Proc. Natl. Acad. Sci. USA* **68**: 2417-2420.
5. ANGELETTI, R. H., R. A. BRADSHAW, AND R. D. WADE. 1971. Subunit structure: Amino acid composition of mouse submaxillary gland nerve growth factor. *Biochemistry* **10**: 463-469.
6. APFEL, S. C., J. C. AREZZO, M. BROWNEE, M. MORAN, P. K. BARNECOTT, AND J. A. KESSLER. 1992. Trophic factor prevention of diabetic neuropathy in a streptozocin rat model. *Soc. Neurosci. Abstr.* **18**: 776.
7. APFEL, S. C., J. C. AREZZO, L. LIPSON, AND J. A. KESSLER. 1992. Nerve growth factor prevents experimental cisplatin neuropathy. *Ann. Neurol.* **31**: 76-80.
8. APFEL, S. C., R. B. LIPTON, J. C. AREZZO, AND J. A. KESSLER. 1991. Nerve growth factor prevents toxic neuropathy in mice. *Ann. Neurol.* **29**: 87-89.
9. ARAKAWA, Y., M. SENDTNER, AND H. THOENEN. 1990. Survival effect of ciliary neurotrophic factor on chick embryonic motoneurons in culture: Comparison with other neurotrophic factors and cytokines. *J. Neurosci.* **10**: 3507-3515.
10. ARMSTRONG, D. M., R. D. TERRY, R. M. N. DETERESA, G. BRUCE, L. B. HERSH, AND F. H. GAGE. 1987. Response of septal

- cholinergic neurons to axotomy. *J. Comp. Neurol.* **264**: 421-436.
11. BABB, T. L., W. R. KUPFER, J. K. PRETORIUS, P. H. CRANDALL, AND M. F. LEVESQUE. 1991. Synaptic reorganization by mossy fibers in human epileptic fascia dentata. *Neuroscience* **42**: 351-363.
 12. BALLARIN, M., P. ERNFORS, N. LINDEFORS, AND H. PERSSON. 1991. Hippocampal damage and kainic acid injection induce a rapid increase in mRNA for BDNF and NGF in the rat brain. *Exp. Neurol.* **114**: 35-43.
 13. BARBIN, G., M. MANTHORPE, AND S. VARON. 1984. Purification of the chick eye ciliary neurotrophic factor. *J. Neurochem.* **43**: 1469-1478.
 14. BARDE, Y.-A., D. EDGAR, AND H. THOENEN. 1982. Purification of a new neurotrophic factor from mammalian brain. *EMBO J.* **1**: 549-553.
 15. BARKER, P. A., C. LOMEN-HOERTH, S. O. MEAKIN, E. M. GENSCHE, AND E. M. SHOOTER. 1992. Trk An: An alternatively spliced form of trk A expressed by neurons. *Soc. Neurosci. Abstr.* **18**: 948.
 16. BARKER, P. A., AND MURPHY, R. A. 1992. The nerve growth factor receptor: A multicomponent system that mediates the actions of the neurotrophin family of proteins. *Mol. Cell. Biochem.* **110**: 1-15.
 17. BECK, K. D., T. H. MCNEILL, C. E. FINCH, F. HEFTI, AND J. R. DAY. 1992. Induction of truncated trk B neurotrophin receptors in hippocampal glial cells during injury-induced axonal sprouting. *Soc. Neurosci. Abstr.* **18**: 920.
 18. BECKWITH, J. B., AND E. V. PERRIN. 1963. *In situ* neuroblastomas: a contribution to the natural history of neural crest tumors. *Am. J. Pathol.* **43**: 1089-1104.
 19. BENEDETTI, M., A. LEVI, AND M. V. CHAO. 1992. Overexpression of the cytoplasmic domain of p75^{NGFR} results in down-regulation of p140^{trk} tyrosine kinase activity. *Soc. Neurosci. Abstr.* **18**: 949.
 20. BENHARROCH, D., AND D. BIRNBAUM. 1990. Biology of the fibroblast growth factor gene family. *Isr. J. Med. Sci.* **26**: 212-219.
 21. BERARDI, N., G. CARMIGNOTO, F. CREMISI, L. DOMENICI, L. MAFFEI, V. PARISI, AND T. PIZZORUSSO. 1990. NGF prevents the change in ocular dominance distribution induced by monocular deprivation in the rat visual cortex. *J. Physiol.* **434**: 14P.
 22. BERG, M. M., D. W. STERNBERG, B. L. HEMPSTEAD, AND M. V. CHAO. 1991. The low-affinity p75 nerve growth factor (NGF) receptor mediates NGF-induced tyrosine phosphorylation. *Proc. Natl. Acad. Sci. USA* **88**: 7106-7110.
 23. BERKEMEIER, L. R., J. W. WINSLOW, D. R. KAPLAN, K. NIKOLICS, D. V. GOEDEL, AND A. ROSENTHAL. 1991. Neurotrophin-5: A novel neurotrophic factor that activates trk and trk B. *Neuron* **7**: 857-866.
 24. BOTHWELL, M. 1991. Keeping track of neurotrophin receptors. *Cell* **65**: 915-918.
 25. BRADLEY, W. G., R. B. DAROFF, G. M. FENICHEL, AND C. D. MARSDEN. 1991. *Neurology in Clinical Practice*. Butterworth-Heinemann, Boston.
 26. CASTREN, E., F. ZAFRA, H. THOENEN, AND D. LINDHOLM. 1992. Light regulates expression of brain-derived neurotrophic factor mRNA in rat visual cortex. *Proc. Natl. Acad. Sci. USA* **89**: 9444-9448.
 27. CATTANEO, E., AND R. MCKAY. 1990. Proliferation and differentiation of neuronal stem cells regulated by nerve growth factor. *Nature* **347**: 762-765.
 28. CHENG, S. V., J. H. NADEAU, R. E. TANZI, P. C. WATKINS, J. JAGADESCH, B. A. TAYLOR, J. L. HAINES, N. SACCHI, AND J. F. GUSELLA. 1988. Comparative mapping of DNA markers from the familial Alzheimer disease and Down syndrome regions of human chromosome 21 to mouse chromosomes 16 and 17. *Proc. Natl. Acad. Sci. USA* **85**: 6032-6036.
 29. COHEN, S., G. CARPENTER, AND L. KING, JR. Epidermal growth factor-receptor protein kinase interactions. *J. Biol. Chem.*, 4834-4842.
 30. COHEN-CORY, S., C. F. DREYFUS, AND I. B. BLACK. 1991. NGF and excitatory neurotransmitters regulate survival and morphogenesis of cultured cerebellar purkinje cells. *J. Neurosci.* **11**: 462-471.
 31. CORDON-CARDO, C., P. TAPLEY, S. JING, V. NANDURI, E. O'ROURKE, F. LAMBALLE, K. KOVARY, R. KLEIN, K. R. JONES, L. F. REICHARDT, AND M. BARBACID. 1991. The trk tyrosine protein kinase mediates the mitogenic properties of nerve growth factor and neurotrophin-3. *Cell* **66**: 173-183.
 32. CORSI, P., AND J. T. COYLE. 1991. Nerve growth factor corrects developmental impairments of basal forebrain cholinergic neurons in the trisomy 16 mouse. *Proc. Natl. Acad. Sci. USA* **88**: 1793-1797.
 33. COULIER, F., D. MARTIN-ZANCA, M. ERNST, AND M. BARBACID. 1989. Mechanism of activation of the human trk oncogene. *Mol. Cell. Biol.* **9**: 15-23.
 34. COYLE, J. T., M. L. OSTER-GRANITE, R. H. REEVES, AND J. D. GEARHART. 1988. Down's syndrome, Alzheimer's disease, and the trisomy 16 mouse. *Trends Neurosci.* **11**: 390-394.
 35. DATE, I., M. F. D. NOTTER, S. Y. FELTEN, AND D. L. FELTEN. 1990. MPTP-treated young mice but not aging mice show partial recovery of the nigrostriatal dopaminergic system by stereotaxic injection of acidic fibroblast growth factor (aFGF). *Brain Res.* **526**: 156-160.
 36. DAVIES, A. M., H. THOENEN, AND Y.-A. BARDE. 1986. Different factors from the central nervous system and periphery regulate the survival of sensory neurones. *Nature* **319**: 497-499.
 37. DAVIES, P., AND A. J. F. MALONEY. 1976. Selective loss of central cholinergic neurons in Alzheimer's disease. *Lancet* **2**: 1403.
 38. DAVIS, S., T. H. ALDRICH, D. M. VALENZUELA, V. V. WONG, M. E. FURTH, S. P. SQUINTO, AND G. D. YANCOPOULOS. 1991. The receptor for ciliary neurotrophic factor. *Science* **253**: 59-63.
 39. DOMENICI, L., N. BERARDI, G. CARMIGNOTO, G. VANTINI, AND L. MAFFEI. 1991. Nerve growth factor prevents the amblyopic effects of monocular deprivation. *Proc. Natl. Acad. Sci. USA* **88**: 8811-8815.
 40. DUNNETT, S. B., AND S. J. RICHARDS. 1990. *Neural Transplantation: From Molecular Basis to Clinical Applications*. Elsevier, Amsterdam.
 41. ENGELE, J., AND M. C. BOHN. 1991. The neurotrophic effects of fibroblast growth factors on dopaminergic neurons *in vitro* are mediated by mesencephalic glia. *J. Neurosci.* **11**: 3070-3078.
 42. ERNFORS, P., J. BENGZON, Z. KOKAIA, H. PERSSON, AND O. LINDVALL. 1991. Increased levels of messenger RNAs for neurotrophic factors in the brain during kindling epileptogenesis. *Neuron* **7**: 165-176.
 43. ERNFORS, P., F. HALLBOOK, T. EBENDAL, E. M. SHOOTER, M. J. RADEKE, T. P. MISKO, AND H. PERSSON. 1988. Developmental and regional expression of β -nerve growth factor receptor mRNA in the chick and rat. *Neuron* **1**: 983-996.
 44. ERNFORS, P., C. F. IBANEZ, T. EBENDAL, L. OLSON, AND H. PERSSON. 1990. Molecular cloning and neurotrophic activities of a protein with structural similarities to nerve growth factor: Developmental and topographical expression in the brain. *Proc. Natl. Acad. Sci. USA* **87**: 5454-5458.
 45. EWING, D. J., AND B. F. CLARKE. 1987. Diabetic autonomic neu-

- ropathy: A clinical viewpoint. In *Diabetic Neuropathy* (P. J. Dyck, T. P. K. Asbury, A. Winegrade, and D. Porte, Jr., Eds.), pp. 66–68. Saunders, Philadelphia.
46. FEINSTEIN, D. L., AND D. LARHAMMER. 1990. Identification of a conserved protein motif in a group of growth factor receptors. *FEBS Lett.* **2**: 7–11.
 47. FERRARI, G., M. C. MINOZZI, G. TOFFANO, A. LEON, AND S. D. SKAPER. 1989. Basic fibroblast growth factor promotes the survival and development of mesencephalic neurons in culture. *Dev. Biol.* **133**: 140–147.
 48. FISCHER, W., F. H. GAGE, AND A. BJORKLUND. 1989. Degenerative changes in forebrain cholinergic nuclei correlate with cognitive impairments in aged rats. *Eur. J. Neurosci.* **1**: 34–45.
 49. FISCHER, W., K. WICTORIN, A. BJORKLUND, L. R. WILLIAMS, S. VARON, AND F. H. GAGE. 1987. Amelioration of cholinergic neuron atrophy and spatial memory impairment in aged rats by nerve growth factor. *Nature* **329**: 65–68.
 50. FREED, W. J., M. POLTORAK, AND J. B. BECKER. 1990. Intracerebral adrenal medulla grafts: A review. *Exp. Neurol.* **110**: 139–166.
 51. FRIEDMAN, W. J., C. F. IBANEZ, F. HALLBOOK, H. PERSSON, L. D. CAIN, C. F. DREYFUS, AND I. B. BLACK. 1992. Differential actions of neurotrophins in the locus coeruleus and basal forebrain. *Soc. Neurosci. Abstr.* **18**: 775.
 52. FUNIBASHI, T., H. SASAKI, AND F. KIMURA. 1988. Intraventricular injection of antiserum to nerve growth factor delays the development of amygdaloid kindling. *Brain Res.* **458**: 132–136.
 53. GAGE, F. H., AND A. BJORKLUND. 1986. Cholinergic septal grafts into the hippocampal formation improve spatial learning and memory in aged rats by a atropine-sensitive mechanism. *J. Neurosci.* **6**: 2837–2847.
 54. GAGE, G. H., D. M. ARMSTRONG, L. R. WILLIAMS, AND S. VARON. 1988. Morphologic response of axotomized septal neurons to nerve growth factor. *J. Comp. Neurol.* **269**: 147–155.
 55. GAGE, F. H., K. WICTORIN, W. FISCHER, L. R. WILLIAMS, S. VARON, AND A. BJORKLUND. 1986. Life and death of cholinergic neurons in the septal and diagonal band region following complete fimbria-fornix transection. *Neuroscience* **19**: 241–255.
 56. GALL, C. M., AND P. J. ISACKSON. 1989. Limbic seizures increase neuronal production of messenger RNA for nerve growth factor. *Science* **245**: 758–761.
 57. GALL, C. M., J. C. LAUTERBORN, AND P. J. ISACKSON. 1990. One paroxysmal discharge stimulates temporally distinct changes in neuronal nerve growth factor and immediate-early gene expression. *J. Cell Biochem. Suppl.* **14F**: 67.
 58. GALL, C., K. MURRAY, AND P. J. ISACKSON. 1991. Kainic acid-induced seizures stimulate increased expression of nerve growth factor mRNA in rat hippocampus. *Mol. Brain Res.* **9**: 113–123.
 59. GARNER, A. S., AND T. H. LARGE. 1992. Evidence for kinaseless and alternate 5' terminal forms of trk B and trk C in chick. *Soc. Neurosci. Abstr.* **18**: 950.
 60. GEINISMAN, Y., F. MORRELL, AND L. DE TOLEDO-MORRELL. 1988. Remodeling of synaptic architecture during hippocampal "kindling." *Proc. Natl. Acad. Sci. USA* **85**: 3260–3264.
 61. GOATE, A., M.-C. CHARTIER-HARLIN, M. MULLAN, J. BROWN, F. CRAWFORD, L. FIDANI, L. GIUFFRA, A. HAYNES, N. IRVING, AND L. JAMES. 1991. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nature* **349**: 704–706.
 62. GOEDERT, M., A. FINE, S. P. HUNT, AND A. ULLRICH. 1986. Nerve growth factor mRNA in peripheral and central rat tissues and in the human central nervous system: Lesion effects in the rat brain and levels in Alzheimer's disease. *Mol. Brain Res.* **1**: 85–92.
 63. GOETZ, C. G., G. T. STEBBINS, H. L. KLAWANS, W. C. KOLLER, R. G. GROSSMAN, R. A. BAKAY, AND R. D. PENN. 1991. United Parkinson Foundation Neurotransplantation Registry on adrenal medullary transplants: Presurgical, and 1- and 2-year follow-up. *Neurology* **41**: 1719–1722.
 64. GOLDE, T. E., S. ESTUS, M. USLAK, L. H. YOUNKIN, AND S. G. YOUNKIN. 1990. Expression of beta amyloid protein precursor mRNAs: Recognition of a novel alternatively spliced form and quantitation in Alzheimer's disease using PCR. *Neuron* **4**: 253–267.
 65. GOTZ, R., F. RAULF, AND M. SCHARTL. 1992. Brain-derived neurotrophic factor is more highly conserved in structure and function than nerve growth factor during vertebrate evolution. *J. Neurochem.* **59**: 432–442.
 66. GREEN, S. H., AND L. A. GREENE. 1986. A single Mr approximately 103,000 ¹²⁵I-beta-nerve growth factor- affinity-labeled species represents both the low and high affinity forms of the nerve growth factor receptor. *J. Biol. Chem.* **261**: 15316–15326.
 67. GREENE, L. A., AND E. M. SHOOTER. 1980. The nerve growth factor: Biochemistry, synthesis, and mechanism of action. *Annu. Rev. Neurosci.* **3**: 352–402.
 68. GREENE, L. A., S. VARON, A. PILTCH, AND E. M. SHOOTER. 1971. Substructure of the β subunit of mouse 7S nerve growth factor. *Neurobiology* **1**: 37–48.
 69. HAGG, T., M. MANTHORPE, V. H. L. VAHLSING, AND S. VARON. 1988. Delayed treatment with nerve growth factor reverses the apparent loss of cholinergic neurons after brain damage. *Exp. Neurol.* **101**: 301–312.
 70. HALLBOOK, F., C. F. IBANEZ, AND H. PERSSON. 1991. Evolutionary studies of the nerve growth factor family reveal a novel member abundantly expressed in Xenopus ovary. *Neuron* **6**: 845–858.
 71. HAROUTUNIAN, V., P. D. KANOF, AND K. L. DAVIS. 1986. Partial reversal of lesion-induced deficits in cortical cholinergic markers by nerve growth factor. *Brain Res.* **386**: 397–399.
 72. HARTIKKA, J., M. STAUFENBIEL, AND H. LUBBERT. 1992. Cyclic AMP, but not basic FGF, increases the *in vitro* survival of mesencephalic dopaminergic neurons and protects the from MPP(+)-induced degeneration. *J. Neurosci. Res.* **32**: 190–201.
 73. HEFTI, F. 1986. Nerve growth factor (NGF) promotes survival of septal cholinergic neurons after fimbria transection. *J. Neurosci.* **6**: 2155–2162.
 74. HEFTI, F., AND D. C. MASH. 1989. Localization of nerve growth factor receptors in the normal human brain and in Alzheimer's disease. *Neurobiol. Aging* **10**: 75–87.
 75. HELGREN, M. E., B. FRIEDMAN, M. KENNEDY, K. MULLHOLLAND, A. MESSER, V. WONG, AND R. M. LINDSAY. 1992. Ciliary neurotrophic factor (CNTF) delays motor impairments in the Mnd mouse, a genetic model of motor neuron disease. *Soc. Neurosci. Abstr.* **18**: 618.
 76. HEMPSTEAD, B. L., D. MARTIN-ZANCA, D. R. KAPLAN, L. F. PARADA, AND M. V. CHAO. 1991. High-affinity NGF binding requires coexpression of the trk proto-oncogene and the low-affinity NGF receptor. *Nature* **350**: 678–682.
 77. HEUMANN, R., S. KORSCHING, J. SCOTT, AND H. THOENEN. 1984. Relationship between levels of nerve growth factor (NGF) and its messenger RNA in sympathetic ganglia and peripheral target tissues. *EMBO J.* **3**: 3183–3189.
 78. HIGASHIJIMA, T. S. UZU, T. NAKAJIMA, AND E. M. ROSS. 1988. Mastoparan, a peptide toxin from wasp venom, mimics receptors by activating GTP-binding regulatory proteins (G proteins). *J. Biol. Chem.* **263**: 6491–6491.

79. HOHN, A., J. LEIBROCK, K. BAILEY, AND Y.-A. BARDE. 1990. Identification and characterization of a novel member of the nerve growth factor/brain-derived neurotrophic factor family. *Nature* **344**: 339-341.
80. HOLMLUND, T. H., H. MITSUMOTO, AND T. GREENE. 1992. The effect of ciliary neurotrophic factor (CNTF) on spontaneously degenerating motor neurons in wobbler mice. *Neurology* **42**(Suppl.):367.
81. HOLTZMAN, D. M., Y. W. LI, S. J. DEARMOND, M. P. MCKINLEY, F. H. GAGE, C. J. EPSTEIN, AND W. C. MOBLEY. 1992. Mouse model of neurodegeneration: atrophy of basal forebrain cholinergic neurons in trisomy 16 transplants. *Proc. Natl. Acad. Sci. USA* **89**: 1383-1387.
82. HOLTZMAN, D. M., Y. LI, L. F. PARADA, S. KINSMAN, C.-K. CHEN, J. S. VALLETTA, J. ZHOU, J. B. LONG, AND W. C. MOBLEY. 1992. p140^{trk} mRNA marks NGF-responsive forebrain neurons: Evidence that *trk* gene expression is induced by NGF. *Neuron* **9**: 465-478.
83. HOSANG, M., AND E. M. SHOOTER. 1985. Molecular characteristics of nerve growth factor receptors on PC12 cells. *J. Biol. Chem.* **260**: 655-662.
84. HURTIG, H., J. JOYCE, J. R. SLADEK, JR., AND J. Q. TROJANOWSKI. 1989. Postmortem analysis of adrenal-medulla-to-caudate autograft in a patient with Parkinson's disease. *Ann. Neurol.* **25**: 607-614.
85. HYMAN, B. T., G. W. VAN HOESEN, AND A. R. DAMASIO. 1990. Memory-related neural systems in Alzheimer's disease: an anatomic study. *Neurology* **40**: 1721-1730.
86. HYMAN, C., M. HOFER, Y.-A. BARDE, M. JUHASZ, G. D. YANCOPOULOS, S. P. SQUINTO, AND R. M. LINDSAY. 1991. BDNF is a neurotrophic factor for dopaminergic neurons of the substantia nigra. *Nature* **350**: 230-232.
87. IBANEZ, C. F., T. EBENDAL, G. BARBARY, J. MURRAY-RUST, T. L. BLUNDELL, AND H. PERSSON. 1992. Disruption of the low affinity receptor-binding site in NGF allows neuronal survival and differentiation by binding to the *trk* gene product. *Cell* **69**: 329-341.
88. IP, N. Y., C. F. IBANEZ, S. H. NYE, J. MCCLAIN, P. F. JONES, D. R. GIES, L. BELLUSCIO, M. M. LE BEAU, R. ESPINOSA, S. P. SQUINTO, H. PERSSON, AND G. D. YANCOPOULOS. 1992. Mammalian neurotrophin-4: structure, chromosomal localization, tissue distribution, and receptor specificity. *Proc. Natl. Acad. Sci. USA* **89**: 3060-3064.
89. ISACKSON, P. J., M. M. HUNTSMAN, K. D. MURRAY, AND C. M. GALL. 1991. BDNF mRNA expression is increased in adult rat forebrain after limbic seizures: temporal patterns of induction distinct from NGF. *Neuron* **6**: 937-948.
90. ITOH, N., S. YONEHARA, A. ISHII, M. YONEHARA, S. MIZUSHIMA, M. SAMESHIMA, A. HASE, Y. SETO, AND S. NAGATA. 1991. The polypeptide encoded by the cDNA for human cell surface antigen Fas can mediate apoptosis. *Cell* **66**: 233-243.
91. JENSEN, L. M. 1987. Phenotypic differentiation of aphidicolin-selected human neuroblastoma cultures after long-term exposure to nerve growth factor. *Dev. Biol.* **120**: 56-64.
92. JING, S., P. TAPLEY, AND M. BARBACID. 1992. Nerve growth factor mediates signal transduction through *trk* homodimer receptors. *Neuron* **9**: 1067-1079.
93. JOHNSON, D., A. LANAHA, C. R. BUC, A. SEHGAL, C. MORGAN, E. MERCER, M. BOTHWELL, AND M. CHAO. 1986. Expression and structure of the human NGF receptor. *Cell* **47**: 545-554, 1986.
94. JOHNSON, E. M., M. TANIUCHI, AND P. S. DISTEFANO. 1988. Expression and possible function of nerve growth factor receptors on Schwann cells. *Trends Neurosci.* **11**: 299-302.
95. JONES, K. R., AND L. F. REICHARDT. 1990. Molecular cloning of a human gene that is a member of the nerve growth factor family. *Proc. Natl. Acad. Sci. USA* **87**: 8060-8064.
96. KAISHO, Y., K. YOSHIURA, AND K. NAKAHAMA. 1990. Cloning and expression of cDNA encoding a novel human neurotrophic factor. *FEBS Lett.* **266**: 187-191.
97. KANG, J., H.-G. LEMAIRE, A. UNTERBECK, J. M. SALBAUM, C. L. MASTERS, K. H. GRZESCHIK, G. MULTHAUP, K. BEYREUTHER, AND B. MULLER-HILL. 1987. The precursor of Alzheimer's disease amyloid A4 protein resembles a cell-surface receptor. *Nature* **325**: 733-736.
98. KAPLAN, D. R., B. L. HEMPSTEAD, D. MARTIN-ZANCA, M. V. CHAO, AND L. F. PARADA. 1991. The *trk* proto-oncogene product: A signal transducing receptor for nerve growth factor. *Science* **252**: 554-560.
99. KAPLAN, D. R., D. MARTIN-ZANCA, AND L. F. PARADA. 1991. Tyrosine phosphorylation and tyrosine kinase activity of the *trk* proto-oncogene product induced by NGF. *Nature* **350**: 158-160.
100. KISS, J., M. SCHLUMPF, AND R. BALAZS. 1989. Selective retardation of the development of the basal forebrain cholinergic and pontine catecholaminergic nuclei in the brain of trisomy 16 mouse, an animal model of Down's syndrome. *Dev. Brain Res.* **50**: 251-264.
101. KITAGUCHI, N., Y. TAKAHASHI, Y. TOKUSHIMA, S. SHIOJIRI, AND H. ITO. 1988. Novel precursor of Alzheimer's disease amyloid protein shows protease inhibitory activity. *Nature* **331**: 530-532.
102. KLEIN, R., D. CONWAY, L. F. PARADA, AND M. BARBACID. 1991. The *trkB* tyrosine protein kinase gene codes for a second neurogenic receptor that lacks the catalytic kinase domain. *Cell* **61**: 647-656.
103. KLEIN, R., S. JING, V. NANDURI, E. O'ROURKE, AND M. BARBACID. 1991. The *trk* proto-oncogene encodes a receptor for nerve growth factor. *Cell* **65**: 189-197.
104. KLEIN, R., D. MARTIN-ZANCA, M. BARBACID, AND L. F. PARADA. 1990. Expression of the tyrosine kinase receptor gene *trkB* is confined to the murine embryonic and adult nervous system. *Development* **109**: 845-850.
105. KNUSEL, B., K. D. BECK, J. W. WINSLOW, A. ROSENTHAL, L. E. BURTON, H. R. WIDMER, K. NIKOLICS, AND F. HEFTI. 1992. Brain-derived neurotrophic factor administration protects basal forebrain cholinergic but not nigral dopaminergic neurons from degenerative changes after axotomy in the adult rat brain. *J. Neurosci.* **12**: 4391-4402.
106. KNUSEL, B., J. W. WINSLOW, A. ROSENTHAL, L. E. BURTON, D. P. SEID, K. NIKOLICS, AND F. HEFTI. 1991. Promotion of central cholinergic and dopaminergic neuron differentiation by brain-derived neurotrophic factor but not neurotrophin-3. *Proc. Natl. Acad. Sci. USA* **88**: 961-965.
107. KOH, S., AND R. LOY. 1988. Age-related loss of nerve growth factor sensitivity in rat basal forebrain neurons. *Brain Res.* **440**: 396-401.
108. KOLIATOS, V. E., H. J. W. NAUTA, R. E. CLATTERBUCK, D. M. HOLZMAN, W. C. MOBLEY, AND D. L. PRICE. 1990. Mouse nerve growth factor prevents degeneration of axotomized basal forebrain cholinergic neurons in the monkey. *J. Neurosci.* **10**: 3801-3813.
109. KONIG, G., C. L. MASTERS, AND K. BEYREUTHER. 1990. Retinoic acid induced differentiated neuroblastoma cells show increased expression of the β A4 amyloid gene of Alzheimer's disease and an altered splicing pattern. *FEBS Lett.* **269**: 305-310.
110. KORSCHING, S., AND H. THONEN. 1983. Nerve growth factor in sympathetic ganglia and corresponding target organs of the

- rat: correlation with density of sympathetic innervation. *Proc. Natl. Acad. Sci. USA* **80**: 3513-3516.
111. KROMER, L. F. 1987. Nerve growth factor treatment after brain injury prevents neuronal death. *Science* **235**: 214-216.
 112. KUMAR, S., J. K. STEWARD, M. WAGHE, D. PEARSON, D. C. EDWARDS, E. L. FENTON, AND A. H. GRIFFITH. 1970. The administration of the nerve growth factor to children with widespread neuroblastoma. *J. Ped. Surg.* **5**: 18-22.
 113. KURZROCK, R., Z. ESTROV, M. WETZLER, J. U. GUTTERMAN, AND M. TALPAZ. 1991. LIF: Not just a leukemia inhibitory factor. *Endo. Rev.* **12**: 208-217.
 114. LAMBALLE, F., R. KLEIN, AND M. BARBACID. 1991. TrkC, a new member of the trk family of tyrosine protein kinases, is a receptor for neurotrophin-3. *Cell* **66**: 967-979.
 115. LANDRETH, G. E., AND E. M. SHOOTER. 1980. Nerve growth factor receptors on PC12 cells: Ligand-induced conversion from low- to high-affinity states. *Proc. Natl. Acad. Sci. USA* **77**: 4751-4755.
 116. LARKFORS, L., T. EBENDAL, S. R. WHITTEMORE, H. PERSSON, B. HOFFER, AND L. OLSON. 1987. Decreased level of nerve growth factor (NGF) and its messenger RNA in the aged rat brain. *Mol. Brain Res.* **3**: 55-60.
 117. LEE, K.-F., E. LI, L. J. HUMBER, S. C. LANDIS, A. H. SHARPE, M. V. CHAO, AND R. JAENISCH. 1992. Targeted mutation of the gene encoding the low affinity NGF receptor p75 leads to deficits in the peripheral sensory nervous system. *Cell* **69**: 737-749.
 118. LEIBROCK, J., F. LOTTSPEICH, A. HOHN, M. HOFER, B. HENGERER, P. MASIAKOWSKI, H. THOENEN, AND Y.-A. BARDE. 1989. Molecular cloning and expression of brain-derived neurotrophic factor. *Nature* **341**: 149-152.
 119. LEVI-MONTALCINI, R. 1987. The nerve growth factor 35 years later. *Science* **237**: 1154-1162.
 120. LEVI-MONTALCINI, R., AND P. U. ANGELETTI. 1968. Nerve growth factor. *Physiol. Rev.* **48**: 534-569.
 121. LILLIEN, L. E., AND P. CLAUDE. 1985. Nerve growth factor is a mitogen for cultured chromaffin cells. *Nature* **317**: 632-634.
 122. LINDSAY, R. M., H. THOENEN, AND Y.-A. BARDE. 1985. Placode and neural crest-derived sensory neurons are responsive at early developmental stages to brain-derived neurotrophic factor. *Dev. Biol.* **112**: 319-328.
 123. LIPTON, R. B., S. C. APFEL, J. P. DUTCHER, R. ROSENBERG, J. KAPLAN, A. BERGER, A. EINZIG, P. WIERNIK, AND H. H. SCHAUMBURG. 1989. Taxol produces a predominantly sensory neuropathy. *Neurology* **39**: 368-373.
 124. LU, B., M. YOKOYAMA, C. F. DREYFUS, AND I. B. BLACK. 1991. Depolarizing stimuli regulate nerve growth factor gene expression in cultured hippocampal neurons. *Proc. Natl. Acad. Sci. USA* **88**: 6289-6292.
 125. McDONALD, N. Q., R. LAPATTO, J. MURRAY-RUST, J. GUNNING, A. WLODAWER, AND T. L. BLUNDELL. 1991. New protein fold revealed by a 2.3 Å resolution crystal structure of nerve growth factor. **354**: 411-414.
 126. MADRAZO, I., R. DURCKER-COLINE, V. DIAZ, J. MARTINEZ-MATA, C. TORRES, AND J. J. BECERRIL. 1987. Open microsurgical autograft of adrenal medulla to the right caudate nucleus in two patients with intractable Parkinsons' disease. *N. Engl. J. Med.* **316**: 831-834.
 127. MAISONPIERRE, P. C., L. BELLUSCIO, S. SQUINTO, N. Y. IP, M. E. FURTH, R. M. LINDSAY, AND G. D. YANCOPOULOS. 1990. Neurotrophin-3: A neurotrophic factor related to NGF and BDNF. *Science* **247**: 1446-1451.
 128. MALLET, S., S. FOSSUM, AND A. N. BARCLAY. 1990. Characterization of the MRC OX40 antigen of activated CD4 positive T lymphocytes—A molecule related to nerve growth factor receptor. *EMBO J.* **9**: 1063-1068.
 129. MARTIN-ZANCA, D., M. BARBACID, AND L. F. PARADA. 1990. Expression of the trk proto-oncogene is restricted to the sensory cranial and spinal ganglia of neural crest origin in mouse development. *Genes Dev.* **4**: 683-694.
 130. MARTIN-ZANCA, D., S. H. HUGHES, AND M. BARBACID. 1986. A human oncogene formed by the fusion of truncated tropomyosin and protein tyrosine kinase sequences. *Nature* **319**: 743-748.
 131. MARTIN-ZANCA, D., R. OSKAM, G. MITRA, T. COPELAND, AND M. BARBACID. 1989. Molecular and biochemical characterization of the human trk proto-oncogene. *Mol. Cell Biol.* **9**: 24-33.
 132. MARUSHIGE, Y., N. R. RAJU, H. MARUSHIGE, AND A. KOESTNER. 1987. Modulation of growth and of morphological characteristics in glioma cells by nerve growth factor and glioma maturation factor. *Cancer Res.* **47**: 4109-4115.
 133. MASLIAH, E., V. LEV-RAM, M. MALLORY, N. GE., M. ELIISMAN, AND T. SAITOH. 1992. The role of amyloid precursor protein in neuritic outgrowth and aberrant sprouting in Alzheimer's disease. *Soc. Neurosci. Abstr.* **18**: 731.
 134. MATSUSHIMA, H., AND E. BOGENMANN. 1990. Nerve growth factor (NGF) induces neuronal differentiation in neuroblastoma cells transfected with the NGF receptor cDNA. *Mol. Cell Biol.* **10**: 5015-5020.
 135. MEAKIN, S. O., AND E. M. SHOOTER. 1991. Tyrosine kinase coupled to the high-affinity NGF receptor complex. *Proc. Natl. Acad. Sci. USA* **88**: 5862-5866.
 136. MEAKIN, S. O., U. SUTER, C. C. DRINKWATER, A. A. WELCHER, AND E. M. SHOOTER. 1991. The rat trk protooncogene product exhibits properties characteristic of the slow nerve growth factor receptor. *Proc. Natl. Acad. Sci. USA* **89**: 2374-2378.
 137. MEDORI, R., L. AUTILIO-GAMBETTI, S. MONACO, AND P. GAMBETTI. 1985. Experimental diabetic neuropathy: Impairment of slow transport with changes in axon cross-sectioned area. *Proc. Natl. Acad. Sci. USA* **82**: 7716-7720.
 138. MIDDLEMAS, D. S., R. A. LINDBERG, AND T. HUNTER. 1991. Trk B, a neural receptor protein-tyrosine kinase: Evidence for a full-length and two truncated receptors. *Mol. Cell Biol.* **11**: 143-153.
 139. MOBLEY, W. C., R. L. NEVE, S. B. PRUSINER, AND M. P. MCKINLEY. 1988. Nerve growth factor increases mRNA levels of the prion protein and the β -amyloid protein precursor in developing hamster brain. *Proc. Natl. Acad. Sci. USA* **85**: 9811-9815.
 140. MOBLEY, W. C., J. L. RUTKOWSKI, G. I. TENNEKON, K. BUCHANAN, AND M. V. JOHNSTON. 1985. Choline acetyltransferase activity in striatum of neonatal rats increased by nerve growth factor. *Science* **229**: 284-287.
 141. MOBLEY, W. C., J. L. RUTKOWSKI, G. I. TENNEKON, J. GEMSKI, K. BUCHANAN, AND M. V. JOHNSTON. 1986. Nerve growth factor increases choline acetyltransferase activity in developing basal forebrain neurons. *Mol. Brain Res.* **1**: 53-62.
 142. MORI, N., S. J. BIRREN, R. STEIN, D. STEMPEL, D. J. VANDENBERGH, C. W. WRENSCHEL, AND D. J. ANDERSON. 1990. Contributions of cell-extrinsic and cell-intrinsic factors to the differentiation of a neural-crest-derived neuroendocrine progenitor cell. *Cold Spring Harb. Symp. Quant. Biol.* **55**: 255-264.
 143. MUFSON, E. J., T. BRASHERS-KRUG, AND J. H. KORDOWER. 1992. p75 Nerve growth factor receptor immunoreactivity in the human brainstem and spinal cord. *Brain Res.* **589**: 115-123.
 144. NAKAGAWARA, A., M. ARIMA, C. G. AZAR, N. J. SCAVARDA, AND G. M. BRODEUR. 1992. Inverse relationship between trk expres-

- sion and n-myc amplification in human neuroblastomas. *Cancer Res.* **52**: 1364-1368.
145. NARUSE, S., S. IGARASHI, H. KOBAYASHI, K. AOKI, T. INUZUKA, K. KANEKO, T. SHIMIZU, K. IIHARA, T. KOJIMA, AND T. MIYATAKE. 1991. Mis-sense mutation Val-Ile in exon 17 of amyloid precursor protein gene in Japanese familial Alzheimer's disease. *Lancet* **337**: 978-979.
 146. NEBREDÁ, A. R., D. MARTIN-ZANCA, D. R. KAPLAN, L. F. PARADA, AND E. SATOS. 1991. Induction by NGF of meiotic maturation of *Xenopus* oocytes expressing the trk proto-oncogene product. *Science* **252**: 558-563.
 147. NEVE, R. L., E. A. FINCH, AND L. R. DAWES. 1988. Expression of the Alzheimer amyloid precursor gene transcripts in the human brain. *Neuron* **1**: 669-677.
 148. OLSON, L., E. O. BACKLUND, T. EBENDAL, R. FREEDMAN, B. HAMBERGER, P. HANSSON, B. HOFFER, U. LINDBLOM, B. MEYERSON, AND I. STROMBERG. 1991. Intraputamin infusion of nerve growth factor to support adrenal medullary autografts in Parkinson's disease. One-year follow-up of first clinical trial. *Arch. Neurol.* **48**: 373-381.
 149. OPPENHEIM, R. W., D. PREVETTE, Y. QIN-WEI, F. COLLINS, AND J. MACDONALD. 1991. Control of embryonic motoneuron survival *in vivo* by ciliary neurotrophic factor. *Science* **251**: 1616-1618.
 150. OPPENHEIM, R. W., Y. QIN-WEI, D. PREVETTE, AND Q. YAN. 1992. Brain-derived neurotrophic factor rescues developing avian motoneurons from cell death. *Nature* **360**: 755-757.
 151. OTTO, D., AND K. UNSICKER. 1990. Basic FGF reverses chemical and morphological deficits in the nigrostriatal system of MPTP-treated mice. *J. Neurosci.* **10**: 1912-1921.
 152. PALLAGE, V., G. TONIOLO, B. WILL, AND F. HEFTI. 1986. Long-term effects of nerve growth factor and neural transplants on behavior of rats with medial-septal lesions. *Brain Res.* **386**: 197-208.
 153. PETERSON, D. I., M. L. PRICE, AND C. S. SMALL. 1989. Autopsy findings in a patient who had an adrenal-to-brain transplant for Parkinson's disease. *Neurol.* **39**: 235-238.
 154. PETERSON, E. R., AND S. M. CRAIN. 1982. Nerve growth factor attenuates neurotoxic effects of taxol on spinal cord-ganglion explants from fetal mice. *Science* **217**: 377-379.
 155. PHILLIPS, H. S., J. N. HAINS, M. ARMANINI, G. R. LARAMÉE, S. A. HONSON, AND J. W. WINSLOW. 1991. BDNF mRNA is decreased in the hippocampus of individuals with Alzheimer's disease. *Neuron* **7**: 695-702.
 156. PIGNATTI, P. F., M. E. BAKER, AND E. M. SHOOTER. 1975. Solution properties of beta nerve growth factor protein and some of its derivatives. *J. Neurochem.* **25**: 155-159.
 157. PIORO, E. P., AND A. C. CUELLO. 1990. Distribution of nerve growth factor receptor-like immunoreactivity in the adult rat central nervous system: Effect of colchicine and correlation with the cholinergic system. II. Brainstem, cerebellum and spinal cord. *Neuroscience* **34**: 89-110.
 158. PONTE, P., P. GONZALEZ-DEQHITT, J. SCHILLING, J. MILLER, D. HSU, B. GREENBERG, K. DAVIS, W. WALLACE, I. LIEBERBURG, F. FULLER, AND B. CORDELL. 1988. A new A4 amyloid mRNA contains a domain homologous to serine proteinase inhibitors. *Nature* **311**: 525-527.
 159. RADEKE, M. J., T. P. MISHO, C. HSU, I. HERZENBERG, AND E. M. SHOOTER. 1987. Gene transfer and molecular cloning of the rat nerve growth factor receptor. *Nature* **325**: 593-597.
 160. RAIVICH, G., A. ZIMMERMAN, AND A. SUTTER. 1985. The spatial and temporal pattern of β -NGF receptor expression in the developing chick embryo. *EMBO J.* **4**: 637-644.
 161. REFOLO, L. M., S. R. J. SALTON, J. P. ANDERSON, P. MEHTA, AND N. K. ROBAKIS. 1989. Nerve and epidermal growth factors induce the release of the Alzheimer amyloid precursor from PC12 cultures. *Biochem. Biophys. Res. Commun.* **164**: 664-670.
 162. RICHARDSON, P. M., V. M. K. VERGE ISSA, AND R. J. RIOPELLE. 1986. Distribution of neuronal receptors for nerve growth factor in the rat. *J. Neurosci.* **6**: 2312-2321.
 163. RODRIGUEZ-TEBAR, A., P. L. JEFFREY, H. THOENEN, AND Y.-A. BARDE. 1989. The survival of chick retinal ganglion cells in response to brain-derived neurotrophic factor depends on their embryonic age. *Dev. Biol.* **136**: 296-303.
 164. ROSENTHAL, A., D. V. GOEDEL, T. NGUYEN, M. LEWIS, A. SHIH, G. R. LARAMÉE, K. NIKOLICS, AND J. W. WINSLOW. Primary structure and biological activity of a novel human neurotrophic factor. *Neuron* **4**: 767-773.
 165. ROSS, A. H., G. SOBUE, H. HOPPA, AND U. R. REDDY. 1991. Biochemical characterization of the nerve growth factor receptor in neural-related tumors. *Curr. Topics Microbiol. Immunol.* **65**: 27-38.
 166. SANDROCK, A. W., JR., AND W. D. MATTHEW. 1987. Substrate-bound nerve growth factor promotes neurite outgrowth in peripheral nerve. *Brain Res.* **425**: 360-363.
 167. SARIOLA, H., M. SAARMA, K. SAINIO, U. ARUMAE, J. PALGI, A. VAAHTOKARI, I. THESLEFF, AND A. KARAVANOV. 1991. Dependence of kidney morphogenesis on the expression of nerve growth factor receptor. *Science* **254**: 571-573.
 168. SATO, Y., P. R. MURPHY, R. SATO, AND H. G. FRIESEN. 1989. Fibroblast growth factor release by bovine endothelial cells and human astrocytoma cells in culture is density dependent. *Mol. Endocrinol.* **3**: 744-748.
 169. SCHECHTER, A. L., AND M. A. BOTHWELL. 1981. Nerve growth factor receptors on PC12 cells: Evidence for two receptor classes with differing cytoskeletal association. *Cell* **24**: 867-874.
 170. SCHECTERSON, L. C., AND M. BOTHWELL. 1992. Novel roles for neurotrophins are suggested by BDNF and NT-3 mRNA expression in developing neurons. *Neuron* **9**: 449-463.
 171. SCHEIBEL, A. B., AND U. TOMIYASU. 1978. Dendritic sprouting in Alzheimer's presenile dementia. *Exp. Neurol.* **60**: 1-8.
 172. SCHIFF, P. B., J. FAUT, AND S. B. HOROWITZ. 1979. Promotion of microtubule assembly *in vitro* by taxol. *Nature* **277**: 665-667.
 173. SCHMIDT, R. E., C. W. MODERT, H. K. YIP, AND E. M. JOHNSON, JR. 1983. Retrograde axonal transport of intravenously administered ¹²⁵I-nerve growth factor in rats with streptozotocin-induced diabetes. *Diabetes* **32**: 654-663.
 174. SCHNEIDER, R., AND M. SCHWEIGER. 1991. A novel modular mosaic of cell adhesion motifs in the extracellular domains of the neurogenic trk and trkB tyrosine kinase receptors. *Oncogene* **6**: 1807-1811.
 175. SCHUBERT, D., L.-W. JIN, T. SITO, AND G. COLE. 1989. The regulation of amyloid β protein precursor secretion and its modulatory role in cell adhesion. *Neuron* **3**: 689-694.
 176. SENDTNER, M., B. HOTMANN, R. KOLBECK, H. THOENEN, AND Y.-A. BARDE. 1992. Brain-derived neurotrophic factor prevents the death of motoneurons in newborn rats after nerve section. *Nature* **360**: 757-759.
 177. SENDTNER, M., G. W. KRETZBER, AND H. THOENEN. 1990. Ciliary neurotrophic factor prevents the degeneration of motor neurons after axotomy. *Nature* **345**: 440-441.
 178. SHELTON, D. L., AND L. F. REICHARDT. 1984. Expression of the β -nerve growth factor gene correlates with the density of sympathetic innervation in effector organs. *Proc. Natl. Acad. Sci. USA* **81**: 7951-7955.

179. SHULTS, C. W. 1992. Future perfect? Presymptomatic diagnosis, neural transplantation, and trophic factors. *Neurol. Clin.* **10**: 567-593.
180. SIEBER-BLUM, M. 1991. Role of neurotrophic factors BDNF and NGF in the commitment of pluripotent neural crest cells. *Neuron* **6**: 949-955.
181. SILANI, V., A. FALINI, O. STRADA, A. PIZZUTI, G. PEZZOLI, E. D. MOTTI, A. VEGETO, AND G. SCARLATO. 1990. Effect of nerve growth factor in adrenal autografts in parkinsonism. *Ann. Neurol.* **27**: 341-342.
182. SLICHENMYER, W. J., AND D. D. VON HOFF. 1990. New natural products in cancer chemotherapy. *J. Clin. Pharmacol.* **30**: 770-788.
183. SLOVITER, R. S. 1992. Possible functional consequences of synaptic reorganization in dentate gyrus of kainate-treated rats. *Neurosci. Lett.* **137**: 91-96.
184. SMITH, C., T. DAVIS, D. ANDERSON, L. SOLAM, M. P. BECKMANN, R. JERZY, S. K. DOWER, D. COSMAN, AND R. G. GOODWIN. 1990. A receptor for tumor necrosis factor defines an unusual family of cellular and viral proteins. *Science* **248**: 1019-1023.
185. SMITH, C. J., D. WION, AND P. BRACHET. 1991. Nerve growth factor-induced neuronal differentiation is accompanied by differential splicing of β -amyloid precursor mRNAs in the PC12 cell line. *Mol. Brain Res.* **10**: 351-354.
186. SOPPET, D., E. ESCANDON, J. MARAGOS, D. S. MIDDLEMAS, S. W. REID, J. BLAIR, L. E. BURTON, B. R. STANTON, D. R. KAPLAN, T. HUNTER, K. NIKOLICS, AND L. F. PARADA. 1991. The neurotrophic factors brain-derived neurotrophic factor and neurotrophin-3 are ligands for the trk B tyrosine kinase receptor. *Cell* **65**: 895-903.
187. SPINA, M. B., S. P. SQUINTO, J. MILLER, R. M. LINDSAY, AND C. HYMAN. 1992. Brain-derived neurotrophic factor protects dopamine neurons against 6-hydroxydopamine and *N*-methyl-4-phenylpyridinium ion toxicity: Involvement of the glutathione system. *J. Neurochem.* **59**: 99-106.
188. SQUINTO, S. P., T. N. STITT, T. H. ALDRICH, S. DAVIS, S. M. BIANCO, C. RADZIEJEWSKI, D. J. GLASS, P. MASIAKOWSKI, M. E. FURTH, D. M. VALENZUELA, P. S. DISTEFANO, AND G. D. YANCOPOULOS. 1991. Trk B encodes a functional receptor for brain-derived neurotrophic factor and neurotrophin-3 but not nerve growth factor. *Cell* **65**: 885-893.
189. STAMENKOVIC, I., E. A. CLARK, AND B. SEED. 1989. A B-lymphocyte activation molecule related to the nerve growth factor receptor and induced by cytokines in carcinomas. *Embo J.* **8**: 1403-1410.
190. STEPHENS, P. H., A. C. CUELLO, M. V. SOFRONIEW, R. C. A. PEARSON, AND P. TAGARI. 1985. The effect of unilateral decortication upon choline acetyltransferase and glutamate decarboxylase activities in the nucleus basalis and other areas of the rat brain. *J. Neurochem.* **45**: 1021-1026.
191. STOCKLI, K. A., F. LOTTSPREICH, M. SENDTNER, P. MASIAKOWSKI, P. CARROLL, R. GOTZ, D. LINDHOLM, AND H. THOENEN. 1989. Molecular cloning, expression and regional distribution of rat ciliary neurotrophic factor. *Nature* **342**: 920-923.
192. STROMBERG, I., M. HERRERA-MARSHITZ, U. UNGERSTEDT, T. EBENDAL, AND L. OLSON. 1985. Chronic implants of chromaffin tissue into the dopamine-denervated striatum: Effects of NGF on graft survival, fiber growth and rotational behavior. *Exp. Brain Res.* **60**: 335-349.
193. SUTTER, A., R. J. RIOPELLE, W. R. M. HARRIS, AND E. M. SHOOTER. 1979. Nerve growth factor receptors: Characterization of two distinct classes of binding sites on chick embryo sensory ganglia cells. *J. Biol. Chem.* **254**: 5972-5982.
194. SUTULA, T., G. CASCINO, J. CAVAZOS, I. PARADA, AND L. RAMIREZ. 1989. Mossy fiber synaptic reorganization in the epileptic human temporal lobe. *Ann. Neurol.* **26**: 321-330.
195. SWEENEY, J. E., C. F. HOHMANN, M. L. OSTER-GRANITE, AND J. T. COYLE. 1989. Neurogenesis of the basal forebrain in euploid and trisomy 16 mice: An animal model for developmental disorders in Down syndrome. *Neuroscience* **31**: 413-425.
196. TAKAHASHI, J. A., M. FUKUMOTO, K. IGARASHI, Y. ODA, H. KIKUCHI, AND M. HATANAKA. 1992. Correlation of basic fibroblast growth factor expression levels with the degree of malignancy and vascularity in human gliomas. *J. Neurosurg.* **76**: 792-798.
197. TANZI, R. E., A. I. MCCLATCHEY, E. D. LAMPERTI, L. VILLAKOMAROFF, J. F. GUSELLA, AND R. NEVE. 1988. Protease inhibitor domain encoded by an amyloid protein precursor mRNA associated with Alzheimer's disease. *Nature* **331**: 528-530.
198. TAUCK, D. L., AND J. V. NADLER. 1985. Evidence of functional mossy fiber sprouting in hippocampal formation of kainic acid-treated rats. *J. Neurosci.* **5**: 1016-1022.
199. TUSZYNSKI, M. H., H. S. U, D. G. AMARAL, AND F. H. GAGE. 1990. Nerve growth factor infusion in the primate brain reduces lesion-induced cholinergic neuronal degeneration. *J. Neurosci.* **10**: 3604-3614.
200. ULLRICH, A., AND J. SCHLESSINGER. 1990. Signal transduction by receptors with tyrosine kinase activity. *Cell* **61**: 203-212.
201. UNSICKER, K., B. DRISCH, J. OTTEN, AND H. THOENEN. 1978. Nerve growth factor-induced fiber outgrowth from isolated rat adrenal chromaffin cells: Impairment by glucocorticoids. *Proc. Natl. Acad. Sci. USA* **75**: 3498-3502.
202. VAN DUJN, C. M., L. HENDRIKS, M. CRUTS, J. A. HARDY, A. HOFMAN, AND C. VAN BROECKHOVEN. 1991. Amyloid precursor protein gene mutation in early-onset Alzheimer's disease. *Lancet* **337**: 978.
203. VINOES, S. A., AND A. KOESTNER. 1981. Effect of nerve growth factor producing cells on anaplastic glioma and pheochromocytoma clones: involvement of other factors. *J. Neurosci. Res.* **6**: 389-401.
204. WESKAMP, G., AND L. F. REICHARDT. 1991. Evidence that biological activity of NGF is mediated through a novel subclass of high affinity receptors. *Neuron* **6**: 649-663.
205. WESTERMANN, R., AND K. UNSICKER. 1990. Basic fibroblast growth factor (bFGF) and rat C6 glioma cells: Regulation of expression, absence of release, and response to exogenous bFGF. *Glia* **3**: 510-521.
206. WHEELER, E. F., AND M. BOTHWELL. 1992. Spatiotemporal patterns of expression of NGF and the low-affinity NGF receptor in rat embryos suggest functional roles in tissue morphogenesis and myogenesis. *J. Neurosci.* **12**: 930-945.
207. WHITE, M. F. 1991. Structure and function of tyrosine kinase receptors. *J. Bioenerg. Biomembr.* **23**: 63-82.
208. WHITEHOUSE, P. J., D. L. PRICE, R. G. STRUBLE, A. W. CLARK, J. T. COYLE, AND M. R. DELONG. 1982. Alzheimer's disease and senile dementia: Loss of neurons in the basal forebrain. *Science* **215**: 1237-1239.
209. WILL, B., AND F. HEFTI. 1985. Behavioral and neurochemical effects of chronic intraventricular injections of nerve growth factor in adult rats with fimbria lesions. *Behav. Brain Res.* **17**: 17-24.
210. WILLIAMS, D. G., AND A. J. WINDEBANK. 1991. Motor neuron

- disease (amyotrophic lateral sclerosis). *Mayo Clin. Proc.* **66**: 54-82.
211. WOODRUFF, N. R., AND K. E. NEET. 1986. Beta nerve growth factor binding to PC12 cells. Association kinetics and cooperative interactions. *Biochemistry* **25**: 7956-7966.
212. WYATT, S., E. M. SHOOTER, AND A. M. DAVIES. 1990. Expression of the NGF receptor gene in sensory neurons and their cutaneous targets prior to and during innervation. *Neuron* **2**: 421-427.
213. YAEGER, M. J., A. KOESTNER, K. MARUSHIGE, AND Y. MARUSHIGE. 1992. The use of nerve growth factor as a reverse transforming agent for the treatment of neurogenic tumors: *In vivo* results. *Acta Neuropathol.* **83**: 624-629.
214. YAN, Q., AND E. M. JOHNSON, JR. 1988. An immunohistochemical study of the nerve growth factor receptor in developing rats. *J. Neurosci.* **8**: 3481-3493.
215. YAN, Q., J. ELLIOTT, AND W. D. SNIDER. 1992. Brain-derived neurotrophic factor rescues spinal motor neurons from axotomy-induced cell death. *Nature* **360**: 753-755.
216. YAN, H., J. SCHLESSINGER, AND M. V. CHAO. 1991. Chimeric NGF-EGF receptors define domains responsible for neuronal differentiation. *Science* **252**: 561-563.
217. YANKNER, B. A., A. CACERES, AND L. K. DUFFY. 1990. Nerve growth factor potentiates the neurotoxicity of β -amyloid. *Proc. Natl. Acad. Sci. USA* **87**: 9020-9023.
218. YOSHIKAWA, K., T. AIZAWA, AND Y. HAYASHI. 1992. Degeneration in vitro of post-mitotic neurons overexpressing the Alzheimer amyloid protein precursor. *Nature* **359**: 64-67.
219. ZAFRA, F., B. HENGERER, J. LEIBROCK, H. THOENEN, AND D. LINDHOLM. 1990. Activity dependent regulation of BDNF and NGF mRNAs in the rat hippocampus is mediated by non-NMDA glutamate receptors. *EMBO J.* **9**: 3545-3550.